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Parenting — a paradigm for investigating the neural circuit basis of behavior Johannes Kohl



Parenting is essential for survival and wellbeing in many species. Since it can be performed with little prior experience and entails considerable sacrifices without immediate benefits for the caregiver, this behavior is likely orchestrated by evolutionarily shaped, hard-wired neural circuits. At the same time, experience, environmental factors and internal state also make parenting highly malleable. These characteristics have made parenting an attractive paradigm for linking complex, naturalistic behavior to its underlying neural mechanisms. Recent work - based on the identification of critical neuronal populations and improved tools for dissecting neural circuits - has uncovered novel functional principles and challenged simplistic models of parenting control. A better understanding of the neural basis of parenting will provide crucial clues to how complex behaviors are organized at the level of cells, circuits and computations. Here I review recent progress, discuss emerging functional principles of parental circuits, and outline future opportunities and challenges.

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Introduction

Building on many decades of research in mammalian model systems, major progress has recently been made in understanding the circuit basis of parental behavior in laboratory mice (*Mus musculus*). Mice are ideally suited to this purpose since they exhibit robust parental care and are genetically tractable. Moreover, powerful tools for circuit mapping and interrogation are available for this species. Neuronal populations crucial for parenting have now been identified and a *functional circuit diagram* underlying parental behavior is taking shape. While these advances have refined previous models and revealed novel principles, they have also uncovered a considerable complexity. Key questions — such as whether parenting relies on dedicated circuits or, rather, generic circuits for social behavior — remain unaddressed. Here I review recent progress, present an emerging circuit logic of parental behavior and outline future challenges. I will first focus on neuronal populations critical for parental behavior before describing an updated functional circuit diagram for parenting. Next, I will discuss the negative regulation of parenting, with novel evidence suggesting that infant-directed aggression is an active process governed by dedicated circuits. Finally, I will outline potential avenues towards a systems-level interrogation of parental behavior.

Neuronal populations critical for parenting

Although strongly modified by experience and physiological state, parenting is an instinctive behavior that can be displayed without any prior experience [1]. For instance, a strain-dependent proportion of virgin female laboratory mice for instance will display spontaneous parental behavior upon first encountering pups, comprising essentially all components of female parental behavior (grooming, licking, crouching, nest building), with the exception of nursing [1]. Similarly, virgin males, in which vomeronasal sensing is abolished, show paternal behavior instead of pup-directed aggression [2,3]. These observations suggest that functional parental circuits are present in adults of both sexes, and that genetic programs strongly contribute to the formation of such circuits. As a consequence, nodes in these circuits are likely composed of defined neuronal populations.

The use of cell type-specific manipulations has considerably advanced our understanding of how parenting as a complex social behavior is organized at the neural level. Most investigations have focussed on brain areas previously identified as critical for parenting by classic lesion studies, such as the medial preoptic area (MPOA) or the posterodorsal medial amygdala (MeApd) [4,5]. Within these areas, neuropeptides, neurotransmitters and receptors have typically been chosen as cellular markers — especially in the hypothalamus, which is composed of a rich set of distinct neuronal cell types [6,7,8[•]]. In addition, immediate early genes (IEGs, e.g. *c-fos*) are frequently used as indirect molecular readouts of neural activity to determine which neurons within such target areas are activated by a given behavior. These approaches have identified parentingrelevant neuronal populations and paved the way for dissecting the circuits within which these neurons function [9^{••},10^{••},11[•],12^{••}]. An initial study from Wu *et al.* reported that MPOA neurons expressing the neuropeptide Galanin (MPOA^{Gal} neurons), which comprise $\sim 20\%$ of MPOA

neurons, are crucial for parental behavior in both sexes (Figure 1) [2]. Two further studies found estrogen receptor α — expressing MPOA neurons (MPOA^{Esr1}) to be critical for pup retrieval in females (Figure 1) [10°,12°]. Intriguingly, MPOA^{Esr1} neurons also strongly affect sexual behavior in males and females [12°].

These observations illustrate several important considerations when using genetic markers for circuit-level studies of behavior: (1) Genetic markers are necessarily imperfect, that is, not all neurons activated by, or involved in controlling, a given behavior, express a single marker. Conversely, not all marker-expressing neurons are involved in a given behavior. Neuropeptide expression can be associated with functional specialization (e.g. somatostatin-positive or parvalbumin-positive interneurons, oxytocinergic and vasopressinergic secretory

Figure 1



Two parenting-relevant neuronal populations in the MPOA. Distribution of, and overlap between, MPOA^{Gal} and MPOA^{Esr1} neurons are shown, as well as the behavioral consequences of manipulating each population. Note that about 90% of MPOA^{Gal} neurons, and more than 80% of MPOA^{Esr1} neurons, are GABAergic [2,12**]. For further details see text. Data from refs. [2,10**,12**] and JK (*unpublished*). Unless specified, manipulations affect behavior in both sexes.

neurons), but such populations are typically involved in narrowly described physiological functions. In contrast, circuits for complex behaviors are unlikely to be defined by single markers. Pragmatic considerations, for example, the availability of Cre mouse lines with restricted expression patterns, seem to underlie marker choice in some cases. (2) In cases where a marker is expressed by the majority of neurons within a brain area (e.g. >50% of MPOA neurons are Esr1-positive (Figure 1) [11[•]] and $\sim 70\%$ of MeApd neurons are GABAergic [13]), the fact that the neurons in question express a marker might be largely irrelevant. Since individual brain areas participate in many behaviors and physiological functions, manipulation of a large fraction of neurons in an area would be expected to result in context-specific effects. This might explain why optogenetic activation of MPOA^{Esr1} neurons elicits context-dependent sexualbehavior or parental-behavior (Figure 1) [12^{••}]. Another prediction is that manipulating variable fractions of a broad population (e.g. by tuning illumination levels in optogenetic experiments) would result in different phenotypes. In cases where the large majority of neurons within an area is manipulated, the conceptual advance over classic, non-cell type specific approaches is questionable. Screening for markers with high *enrichment ratios*, that is, controlling for relative frequency of markerpositive neurons within an area can address this limitation (see [2]). (3) Immediate early genes such as *c*-fos are slow (minutes-hours) and only provide an indirect readout of neural activity. Also, it remains incompletely understood which neuronal activity patterns result in their activation in vivo [14]. IEG-positive and marker-positive neurons thus only partially reflect parenting-relevant neural populations. These limitations also apply to other systems, such as Esr1-expressing neurons in the ventrolateral ventromedial nucleus of the hypothalamus (VMHvl^{Esr1}), which have prominent roles in aggression [15] but also food intake, physical activity and thermogenesis [16,17].

Single-cell and spatial transcriptomics approaches now offer the opportunity to further define neuronal populations based on location, anatomical connectivity and gene expression profile [8,18-22]. Several recent studies have used such approaches on hypothalamic populations $[6,8^{\circ}]$. For instance, Moffitt et al. recently assembled a spatially resolved molecular atlas of the MPOA, identifying distinct MPOA^{Gal} subpopulations [8[•]]. In order to functionally exploit such refined molecular identities, better genetic access to such neuronal populations is required. At present, neurons characterized by expressing single marker genes are typically targeted using recombinaseexpressing mouse lines. Only a handful of orthogonal recombinases (Cre, Flp, Dre, Φ C31, Vika) are currently available [23-26]. Of those, Cre accounts for the vast majority and the generation of new lines is slow and expensive. Genetic intersections therefore remain challenging and impractical. Alternatively, conditional viral tools, especially adeno-associated viruses (AAVs), can be used. While their limited packaging capacity (~4.7 kb) often precludes the incorporation of promoter fragments large enough to drive cell-type specific transgene expression (but see e.g. [27,28]), enhancer sequences have been shown to be suitable for this purpose [27,29]. Such approaches have the potential to give access to more specific, behaviorally relevant neuronal populations in the future.

Circuit logic of parenting

Behaviors are encoded by dynamic activity patterns in brain-wide circuits. Although specific neuronal populations can neither be necessary nor sufficient for any given behavior [30], the identification of parenting-relevant neuronal populations has recently precipitated rapid advances in our understanding of how parenting is orchestrated at the circuit level [9^{••},12^{••},31,32[•]]. Lesion studies and pharmacological manipulations, primarily in female rats, have found many brain areas to be involved in parenting [1,9^{••},33,34]. Importantly, each of these areas is also critical for other social and non-social behaviors. Based on these seminal studies, a circuit model for parenting was proposed in which two opposing pathways mediate the activation and inhibition of parenting, respectively [1]. Chemosensory pup stimuli are integrated by the MeA, which exerts a negative effect on parenting by directly inhibiting the MPOA and by activating a 'central aversion network', encompassing the (ventral) lateral septum (LS), anterior hypothalamic nucleus (AH), VMH, dorsal premammillary nucleus (PMd) and periaqueductal gray (PAG). In contrast, the MPOA and adjacent ventral bed nucleus of the stria terminalis (vBNST) promote parenting, controlling its distinct components via dedicated downstream projections [1].

Recent work in mice has begun to develop this regionlevel wiring diagram (lacking cellular identity and signs of synaptic connections) into a functional circuit diagram (Figure 2), starting from genetically defined populations such as MPOA^{Gal} neurons. Conditional retrograde transsynaptic and anterograde viral tracers have been used to anatomically delineate elements of the circuit in which MPOA^{Gal} neurons are embedded [9^{••}]. These neurons project to, and receive inputs from, more than 20 brain areas in a circuit exhibiting extensive reciprocity [9^{••}]. Importantly, MPOA^{Gal} neurons form projection-defined subpopulations, each receiving inputs from essentially all input areas (Figure 2) [9**]. The parallel organization of MPOA^{Gal} projections is similar to what has been described for agouti-related peptide-expressing neurons in the arcuate nucleus (ArcAgrp neurons) [35], but contrasts with, for example, VMH^{Esr1} or PeFA^{Ucn3} neurons (see 'Negative regulation of parenting'), which predominantly send out branched projections [36,37]. Corresponding with this segregated organization,

different MPOA^{Gal} pools are active during different episodes of parenting, and control distinct motor, motivational and hormonal aspects of parenting (Figure 2) [9^{••}]. For instance, projections to the periaqueductal gray (PAG) are critical for pup grooming, which recapitulates the effect of optogenetically activating the entire MPOA-^{Gal} population [2]. In contrast, MPOA^{Gal} projections to the ventral tegmental area (VTA) seem to control the motivation to interact with pups [9^{••}]. In a separate study, Fang et al. reported that stimulating VTA-projecting MPOA^{Esr1} neurons elicits pup retrieval to the nest [10^{••}], identical to what is observed when *all* Esr1expressing or GABA-expressing MPOA neurons are activated [11[•],12^{••}]. VTA-mediated pup retrieval might be a consequence of acutely increased parental motivation (stimulation of MPOA^{Esr1} neurons also elicits retrieval of rubber pups [12^{••}]), but further experimental evidence is needed to address the role of this projection. While it remains to be shown whether these projectiondefined MPOA subpopulations have separable genetic identities (see e.g. [8]), these results indicate that discrete components of a complex behavior can be isolated at the circuit level.

In addition to such efforts to trace parenting-relevant circuits in an *inside-out* manner, i.e. starting from neuronal populations deep in the brain, another possibility is to define parental circuits in an *outside-in* manner, starting from the sensory periphery. Such efforts have encountered both methodological and conceptual hurdles. One technical challenge is the absence of suitable reagents for anterograde trans-synaptic circuit tracing, although progress has recently been made in this regard [38]. Other limitations are of a conceptual nature: Because of their presumed ability to 'trigger' instinctive behaviors, pheromonal cues have long been proposed to be processed along dedicated, stimulus-specific neural circuits from the sensory periphery into the brain (labeled lines) [39]. Pup-emitted pheromones are thought to promote pup-directed aggression, since ablating vomeronasal organ (VNO) function elicits paternal behavior in otherwise infanticidal virgin males [2,3]. The identification of pup-specific vomeronasal receptors (VRs) might therefore constitute entry points into labeled line circuits into the brain. However, a recent study found that neither pup-sensitive vomeronasal receptors nor associated cues are pup-specific [40^{••}]. Instead, such receptors are also tuned to adult chemosensory signals, and pup recognition relies on a combination of physical and chemical traits (see 'Negative regulation of parenting') [40^{••}]. These findings thus call into question the existence of labeled lines for pheromone-triggered behavior [39,41], and therefore the possibility of an *outside-in* identification of parental circuits.

In summary, considerable progress has been made in uncovering the functional circuit architecture underlying



Figure 2

Emerging circuit logic underlying parental behavior. This functional circuit diagram is based on pharmacological and lesion- studies in virgin female rats [1], and extended by recent findings (see text, refs. [9^{••},10^{••},11[•],12^{••},32[•],37,40^{••},42,53^{••}]). Arc^{Agrp} neurons, which sense caloric need and mediate feeding behavior, project to a subset of MPOA neurons [11[•]]. Optogenetic stimulation of this projection decreases maternal nestbuilding [11[•]]. Tyrosin hydroxylase-expressing neurons in the anteroventral periventricular nucleus (AVPeTH neurons) are critical for parental behavior in females [42]. These neurons form monosynaptic connections with oxytocin-expressing neurons in the paraventricular hypothalamic nucleus, thereby influencing oxytocin release [42]. Abbreviations: AHI, amygdalohippocampal area; AOB, accessory olfactory bulb; AVPe, anteroventral periventricular nucleus; BNST, bed nucleus of the stria terminalis; LC, locus coeruleus; LS, lateral septum; IHb, lateral habenula; MeA, medial amygdala; NAc, nucleus accumbens; PVN, periventricular hypothalamic nucleus; PVT, periventricular thalamic nucleus; RRF, retrorubral field; SNpc, substantia nigra pars compacta; somat ctx, somatosensory cortex; SON, supraoptic nucleus; Vglut, vesicular glutamate transporter; Vgat, vesicular GABA transporter; VMH, ventromedial hypothalamus; VTA, ventral tegmental area.

parental behavior. Key emerging principles are that these circuits are enormously complex, overall remarkably similar between the sexes (but see [32°,42]), and that specific aspects of parenting can indeed be assigned to discrete circuit elements [9°,43]. It will be interesting to investigate how this circuitry interacts with neural systems controlling other instinctive behaviors (or whether they largely overlap), how information is

processed between successive circuit nodes and how experience and physiological states affect their function.

Negative regulation of parenting

Under certain physiological and environmental conditions, animals neglect or attack young conspecifics. Males in some species kill unfamiliar infants to gain reproductive advantage [44–46] and females neglect or

attack their young during stressful circumstances such as food shortage or threat of predation [47]. In laboratory mouse strains, which are a product of artificial selection for a variety of physiological and behavioral traits [45,47,48], aggressive behavior is less pronounced. Infant-directed aggression has predominantly been studied in males, which undergo a switch from infanticide to paternal care after mating [2,3,49,50]. This striking phenomenon seems to be synchronized with the female's gestation time to ensure paternity. Does infanticide simply result from downregulating parental circuits, or is it rather orchestrated by dedicated circuits? Several lines of evidence now indicate that it is a combination of both, as I will outline below.

In the periphery, detection of pup-emitted chemosensory signals is crucial for male infanticide, since this behavior is abolished by surgical or genetic ablation of VNO function [2,3]. Two recent studies have identified relevant cues and neurons involved in their detection: Trouillet et al. found that conditional ablation of the G-protein subunit Gai2 — expressed in a subclass of VNO neurons — reduces infant-directed aggression. In a complementary study, Isogai et al. systematically screened for VNO neurons (which each typically express a single VR) activated by pup cues. They identified a repertoire of 7 VRs, knock-out of two of which, Vmn2r65 and Vmn2r88 (both Gai2-negative), significantly decreased pup-directed aggression in virgin males [40^{••}]. Together, these results suggest that several VRs (and, correspondingly, VNO neuron types) contribute to the detection of infant cues. Surprisingly, however, these VRs are also activated by adult cues, and pup recognition requires a combination of chemical and tactile cues [40^{••}]. Furthermore, the chemical stimuli detected by Vmn2r65 and Vmn2r88 are rather unexpected: submandibular gland protein C, expressed in salivary glands of pups and adult females, and hemoglobins, which are ubiquitously found in social environments, especially after parturition [40**]. These results indicate that VNO cues emitted by infants are ambiguous, and that adults use multisensory information for pup recognition.

How are pro-infanticidal stimuli processed deeper in the brain? Vomeronasal information is relayed to the MeA via the accessory olfactory bulb (AOB) before reaching hypothalamic areas, such as the BNST or MPOA (Figure 2) [51]. Chemosensory signals from both VNO and the main olfactory system are presumably integrated by MeA neurons [52], but it remains unclear where and how these signals interact with haptic and other types of sensory information to form pup representations (Figure 2). Intriguingly, ablation of G α i2 suppresses infanticide, but enhances male-male aggression [53^{••}]. Together with the observation that the MeA neurons activated during infanticide are different from those

involved in male-male aggression [53^{••}], this suggests that these aggressive behaviors are controlled by different circuit mechanisms. MeA lesions facilitate parental behavior in females, and activation of GABAergic MeA neurons mirrors this effect [32°,54]. The effects of MeA lesions on pup-directed behavior in males are unclear, but Chen et al. recently reported that optogenetic activation of GABAergic MeA neurons can result in either parental behavior or infanticide, depending on illumination strength [32[•]]. Since the large majority of MeA neurons are GABAergic [55], these effects might be the consequence of activating neuronal subpopulations with distinct roles (see 'Neuronal populations critical for parenting'). Located further along the pheromone processing pathway, lesions to the rhomboid nucleus of the BNST (BSTrh) were shown to suppress infanticidal behavior [56[•]], and functional maturation of BSTrh inputs during adolescence has been hypothesized to underlie the change from parental to infanticidal behavior [57[•]]. In order to identify additional infanticide-relevant regions, a recent study used brain-wide IEG mapping, uncovering a marked upregulation of c-Fos in the caudal hypothalamus after pup-directed aggression [58]. Autry et al. subsequently investigated this region in greater detail and found that Urocortin 3-expressing neurons in the perifornical area (PeFA^{Ucn3} neurons) are activated during pup-directed, but not male-male, aggression in both sexes [37]. While silencing of PeFA^{Ucn3} neuronal activity in virgin males blocks infanticide, activation of these neurons elicits infant-directed neglect in virgin females [37]. Intriguingly, PeFA^{Ucn3} neurons receive direct inputs from (almost exclusively inhibitory) MPOA^{Gal} neurons [2], suggesting that infanticide-promoting circuits might be actively suppressed in parental animals.

Altogether, these observations indicate that (1) infantdirected aggression relies on dedicated circuits which are likely distinct from those mediating male-male aggression, (2) these circuits directly interact with parental circuits — potentially in a mutually inhibitory fashion, and (3) similar neural mechanisms control infant-directed aggression in males and females. It will be exciting to further dissect the circuit mechanisms underlying infantdirected aggression, to investigate how stress promotes this behavior in females, and to address which plasticity mechanisms govern the switch from infanticide to parenting in males.

Towards a systems-level investigation of parental behavior

A key insight from recent studies is that parenting, as well as other instinctive behaviors, rely on highly complex, unexpectedly malleable, and potentially overlapping circuits [9**,35,59,60]. It remains unclear whether parental behavior is controlled by parenting-specific circuits or rather by general-purpose social behavior circuits that are state-specifically and/or context-specifically engaged. Distinguishing between these scenarios will require the use of systems neuroscience approaches and the integration of anatomical, functional and behavioral data.

First, single cell and spatial transcriptomics approaches have the potential to identify novel genetic entry points into parenting-relevant neuronal populations, and to uncover plasticity mechanisms within these populations. For instance. Moffitt *et al.* recently used a massively multiplexed in situ hybridization pipeline (MERFISH) to create a cell atlas of the preoptic area, defining novel cell types and subdividing MPOA^{Gal} neurons into ten transcriptionally and spatially distinct clusters [8[•]]. Second, refined anatomical approaches will help uncover further motifs in parental circuits, thereby guiding future functional investigations. Improved viral vectors now enable more specific, efficient and permanent access to defined neurons and circuits [61-65]. However, viral tracing approaches typically visualize connectivity between hundreds to thousands of neurons, thereby obscuring cellular-level anatomical diversity. Individual neurons can be reconstructed by serial two-photon tomography after sparse neuronal labeling, which revealed strikingly complex morphologies and brain-wide projection patterns [66,67]. However, this approach is is highly time-consuming, resource-intensive and laborious. High-throughput, sequencing-based strategies, such as MapSeq [68] are expected to give complementary insights into the organizational principles of parentingrelevant circuits. Third, rather than investigating these circuits one node at a time, addressing dynamic information processing at brain-wide scales will be necessary to understand the neural computations underlying parenting and other instinctive behaviors. High-density recordings from thousands of individually resolved neurons across the brain, will be instrumental for tracking information flow within circuits [69-72]. Lastly, deep learning approaches now allow for automated, markerless tracking of animals under varying experimental conditions, thereby greatly reducing the time required to analyze behavioral video recordings [73-75]. These methods have facilitated behavioral tracking, but behavioral *classification* remains challenging (e.g. pup grooming versus chemoinvestigation), especially for social interactions involving several subjects. Further improvements to these algorithms, assay-specific behavioral classifiers, and optimization of experimental conditions will without doubt result in increasingly automated behavioral quantification.

Fully leveraging these methodologies will put us in a position to address key questions in neuroscience, such as the degree of plasticity within neural circuits thought to be hardwired, how robustness and plasticity are balanced in such systems, and whether circuits for different behaviors are separate or highly overlapping. Thus, insights into the neural mechanisms underlying parental behavior have the potential to broadly contribute to our general understanding of how evolutionarily sculpted circuits control instinctive behaviors.

Conflict of interest statement

Nothing declared.

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References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- •• of outstanding interest
- 1. Numan M, Insel TR: *The Neurobiology of Parental Behavior*. Springer; 2003.
- Wu Z, Autry AE, Bergan JF, Watabe-Uchida M, Dulac CG: Galanin neurons in the medial preoptic area govern parental behaviour. Nature 2014, 509:325-330.
- Tachikawa KS, Yoshihara Y, Kuroda KO: Behavioral Transition from Attack to Parenting in Male Mice: A Crucial Role of the Vomeronasal System. 2013 http://dx.doi.org/10.1523/ JNEUROSCI.2364-12.2013.
- Numan M: Medial preoptic area and maternal behavior in the female rat. J Comp Physiol Psychol 1974, 87:746-759.
- Numan M, Numan MJ, English JB: Excitotoxic amino acid injections into the medial amygdala facilitate maternal behavior in virgin female rats. *Horm Behav* 1993, 27:56-81.
- Romanov RA, Zeisel A, Bakker J, Girach F, Hellysaz A, Tomer R, Alpár A, Mulder J, Clotman F, Keimpema E et al.: Molecular interrogation of hypothalamic organization reveals distinct dopamine neuronal subtypes. Nat Neurosci 2017, 20:176-188.
- Romanov RA, Alpár A, Hökfelt T, Harkany T: Unified classification of molecular, network, and endocrine features of hypothalamic neurons. *Annu Rev Neurosci* 2019, 42:1-26.
- 8. Moffitt JR, Bambah-Mukku D, Eichhorn SW, Vaughn E, Shekhar K,
- Perez JD, Rubinstein ND, Hao J, Regev A, Dulac Cet al.: Molecular, spatial, and functional single-cell profiling of the hypothalamic preoptic region. Science 2018, 362(6416) eaau5324

The authors use single-cell transcriptomics and spatial transcriptomics to build a high-resolution cell atlas of the preoptic area (POA) of the hypothalamus. Combining spatial expression profiling with immediate early gene (IEG) labelling, the authors identify POA neuron types activated by specific social behaviors such as parenting, aggression and mating.

- 9. Kohl J, Babayan BM, Rubinstein ND, Autry AE, Marin-Rodriguez B,
- Kapoor V, Miyamishi K, Zweifel LS, Luo L, Uchida N et al.: Functional circuit architecture underlying parental behaviour. Nature 2018, 556:326-331

This study finds that different projection-defined subpopulations of Galanin-expressing neurons in the medial preoptic area mediate distinct aspects of parental behavior in both sexes.

- 10. Fang YY, Yamaguchi T, Song SC, Tritsch NX, Lin D: A
- hypothalamic midbrain pathway essential for driving maternal behaviors. Neuron 2018, 98(1):192-207
 Inactivation of MPOA^{Esr1} neurons impairs pup approach and retrieval in

Inactivation of MPOA^{Esr1} neurons impairs pup approach and retrieval in females, whereas optogenetic activation induces immediate pup retrieval. Inhibitory projections from MPOA^{Esr1} neurons to predominantly non-dopaminergic neurons in the ventral tegmental area (VTA) are essential for driving pup retrieval, and VTA dopaminergic cells are reliably activated during this behavior.

- Li X-Y, Han Y, Zhang W, Wang S-R, Wei Y-C, Li S-S, Lin J-K, Yan J J, Chen A-X, Zhang X et al.: Agrp neurons project to the medial preoptic area and modulate maternal nest-building. J Neurosci 2019, 39(3):456-471

Li et al. demonstrate that Arc^{Agrp} neurons form direct, inhibitory projections onto a subset of MPOA neurons. Optogenetic activation of this projection decreases maternal nest building, but only minimally affects pup retrieval. This study therefore provides a possible circuit mechanism by which food restriction could suppress aspects of parental behavior.

- 12. Wei Y-C, Wang S-R, Jiao Z-L, Zhang W, Lin J-K, Li X-Y, Li S-S,
- Zhang X, Xu X-H: Medial preoptic area in mice is capable of ••
- mediating sexually dimorphic behaviors regardless of gender. Nat Commun 2018, 9:279 Optogenetic activation of MPOA neurons elicits context-dependent

mounting and pup retrieval in both males and females, hinting at sexshared neural architecture for these behaviors. Esr1-expressing MPOA neurons are essential for the sexually biased display of these behaviors.

- 13. Li Y, Mathis A, Grewe BF, Osterhout JA, Ahanonu B, Schnitzer MJ, Murthy VN, Dulac C: Neuronal representation of social information in the medial amygdala of awake behaving mice. Cell 2017, 171(5):1176-1190.
- 14. Mahringer D. Petersen AV. Fiser A. Okuno H. Bito H. Perrier J-F. Keller GB: Expression of c-Fos and Arc in hippocampal region CA1 marks neurons that exhibit learning-related activity changes. bioRxiv 2019:644526.
- 15. Lee H, Kim D-W, Remedios R, Anthony TE, Chang A, Madisen L, Zeng H, Anderson DJ: Scalable control of mounting and attack by Esr1+ neurons in the ventromedial hypothalamus. Nature 2014. 509:627-632
- 16. Choi Y-H, Fujikawa T, Lee J, Reuter A, Kim KW: Revisiting the ventral medial nucleus of the hypothalamus: the roles of SF-1 neurons in energy homeostasis. Front Neurosci 2013, 7:71.
- 17. Correa SM, Newstrom DW, Rubenstein JL, Ingraham HA: An estrogen-responsive module in the ventromedial hypothalamus selectively drives sex-specific activity in females. Cell Rep 2015, 10:62-74.
- 18. Ståhl PL, Salmén F, Vickovic S, Lundmark A, Navarro JF, Magnusson J, Giacomello S, Asp M, Westholm JO, Huss M et al.: Visualization and analysis of gene expression in tissue sections by spatial transcriptomics. Science (80-) 2016, 353:78-82.
- Vickovic S, Eraslan G, Salmén F, Klughammer J, Stenbeck L, 19. Äijö T, Bonneau R, Bergenstråhle L, Navarro JF, Gould J et al .: High-density spatial transcriptomics arrays for in situ tissue profiling. bioRxiv 2019:563338.
- 20. Eng C-HL, Lawson M, Zhu Q, Dries R, Koulena N, Takei Y, Yun J, Cronin C, Karp C, Yuan G-C *et al.*: Transcriptome-scale superresolved imaging in tissues by RNA seqFISH+. Nature 2019, 568·235-239
- 21. Rodriques SG, Stickels RR, Goeva A, Martin CA, Murray E, Vanderburg CR, Welch J, Chen LM, Chen F, Macosko EZ: Slideseq: a scalable technology for measuring genome-wide expression at high spatial resolution. Science (80-) 2019, 363:1463-1467.
- 22. Wang X, Allen WE, Wright MA, Sylwestrak EL, Samusik N, Vesuna S, Evans K, Liu C, Ramakrishnan C, Liu J et al.: Threedimensional intact-tissue sequencing of single-cell transcriptional states. Science 2018, 361:eaat5691.
- 23. Karimova M, Baker O, Camgoz A, Naumann R, Buchholz F, Anastassiadis K: A single reporter mouse line for Vika, Flp, Dre, and Cre-recombination. Sci Rep 2018, 8:14453.
- 24. Madisen L, Garner AR, Shimaoka D, Chuong AS, Klapoetke NC, Li L, van der Bourg A, Niino Y, Egolf L, Monetti C et al.: Transgenic mice for intersectional targeting of neural sensors and effectors with high specificity and performance. Neuron 2015, 85:942-958
- Anastassiadis K, Fu J, Patsch C, Hu S, Weidlich S, Duerschke K, Buchholz F, Edenhofer F, Stewart AF: Dre recombinase, like Cre, 25. is a highly efficient site-specific recombinase in E. coli, mammalian cells and mice. Dis Model Mech 2009, 2:508-515
- 26. Imayoshi I, Hirano K, Kitano S, Miyachi H, Kageyama R: In vivo evaluation of PhiC31 recombinase activity in transgenic mice Neurosci Res 2012, 73:106-114.

- 27. Jüttner J, Szabo A, Gross-Scherf B, Morikawa RK, Rompani SB, Hantz P, Szikra T, Esposti F, Cowan CS, Bharioke A et al.: Targeting neuronal and glial cell types with synthetic promoter AAVs in mice, non-human primates and humans. Nat Neurosci 2019. 22:1345-1356.
- 28. Fields RL, Ponzio TA, Kawasaki M, Gainer H: Cell-type specific oxytocin gene expression from AAV delivered promoter deletion constructs into the rat supraoptic nucleus in vivo. PLoS One 2012. 7:e32085.
- 29. Graybuck LT, Sedeño-Cortés A, Nguyen TN, Walker M, Szelenyi E, Lenz G, Sieverts L, Kim TK, Garren E, Kalmbach B et al.: Prospective, brain-wide labeling of neuronal subclasses with enhancer-driven AAVs. bioRxiv 2019:525015.
- Yoshihara M, Yoshihara M: 'Necessary and sufficient' in biology 30. is not necessarily necessary – confusions and erroneous conclusions resulting from misapplied logic in the field of biology, especially neuroscience. J Neurogenet 2018, 32:53-64.
- 31. Lin R, Li Y, Luo M: A neural circuit driving maternal behaviors. Neuron 2018, 98(1):6-8.
- 32. Chen PB, Hu RK, Wu YE, Pan L, Huang S, Micevych PE, Hong W: Sexually dimorphic control of parenting behavior by the medial amygdala. Cell 2019, 176:1206-1221.e18

GABAergic - but not glutamatergic - neurons in the medial amygdala (MeA) promote parental behavior in females, whereas in males, scalable activation of GABAergic MeA neurons elicits both parenting and infanticide.

- 33. Dulac C, O'Connell LA, Wu Z: Neural control of maternal and paternal behaviors. Science (80-) 2014, 345:765-770.
- 34. Zilkha N, Scott N, Kimchi T: Sexual dimorphism of parental care: from genes to behavior. Annu Rev Neurosci 2017, 40:273-305.
- 35. Betley JN, Cao ZFH, Ritola KD, Sternson SM: Parallel, redundant circuit organization for homeostatic control of feeding behavior. Cell 2013, 155:1337-1350.
- 36. Lo L, Yao S, Kim D-W, Cetin A, Harris J, Zeng H, Anderson DJ, Weissbourd B: Connectional architecture of a mouse hypothalamic circuit node controlling social behavior. Proc Natl Acad Sci U S A 2019, 116:7503-7512.
- Autry AE, Wu Z, Kohl J, Bambah-Mukku D, Rubinstein ND, Marin-Rodriguez B, Carta I, Sedwick V, Dulac C: Perifornical area urocortin-3 neurons promote infant-directed neglect and aggression. bioRxiv 2019:697334.
- Zeng W-B, Jiang H-F, Gang Y-D, Song Y-G, Shen Z-Z, Yang H, Dong X, Tian Y-L, Ni R-J, Liu Y et al.: Anterograde monosynaptic transneuronal tracers derived from herpes simplex virus 1 strain H129. Mol Neurodegener 2017, 12:38.
- Ishii KK, Osakada T, Mori H, Miyasaka N, Yoshihara Y, Miyamichi K, Touhara K: A labeled-line neural circuit for pheromone-mediated sexual behaviors in mice. Neuron 2017, 95:123-137.e8
- 40. Isogai Y, Wu Z, Love MI, Ahn MHY, Bambah-Mukku D, Hua V,
 Farrell K, Dulac C: Multisensory logic of infant-directed aggression by males. Cell 2018, 175(7):1827-1841

Isogai et al. show that pup salivary chemosensory signals combined with morphological features are important to elicit male aggression. This suggests that infant recognition relies on integration of multiple sensory modalities.

- Osakada T, Ishii KK, Mori H, Eguchi R, Ferrero DM, Yoshihara Y, Liberles SD, Miyamichi K, Touhara K: Sexual rejection via a vomeronasal receptor-triggered limbic circuit. Nat Commun 2018, 9:4463.
- 42. Scott N, Prigge M, Yizhar O, Kimchi T: A sexually dimorphic hypothalamic circuit controls maternal care and oxytocin secretion. Nature 2015, 525(7570):519-523.
- 43. Horrell ND, Perea-Rodriguez JP, Harris BN, Saltzman W: Effects of repeated pup exposure on behavioral, neural, and adrenocortical responses to pups in male California mice (Peromyscus californicus). Horm Behav 2017, 90:56-63.
- 44. Hrdy SB: Infanticide among animals: a review, classification, and examination of the implications for the reproductive strategies of females. Ethol Sociobiol 1979, 1:13-40.

- 45. Lukas D, Huchard E: The evolution of infanticide by males in mammalian societies. *Science* (80-) 2014, **346**:841-844.
- 46. Parmigiani S, Saal FS, vom, Saal FS, vom: *Infanticide and Parental Care*. Routledge; 2016.
- 47. Lukas D, Huchard E: The evolution of infanticide by females in mammals. *Philos Trans R Soc B Biol Sci* 2019, **374**:20180075.
- 48. Svare B, Bartke A: Food deprivation induces conspecific pupkilling in mice. *Aggress Behav* 1978, 4:253-261.
- Vom Saal FS: Time-contingent change in infanticide and parental behavior induced by ejaculation in male mice. *Physiol Behav* 1985, 34:7-15.
- Labov JB: Factors influencing infanticidal behavior in wild male house mice (*Mus musculus*). Behav Ecol Sociobiol 1980, 6:297-303.
- 51. Dulac C, Wagner S: Genetic analysis of brain circuits underlying pheromone signaling. Annu Rev Genet 2006, 40:449-467.
- Keshavarzi S, Power JM, Albers EHH, Sullivan RKS, Sah P: Dendritic organization of olfactory inputs to medial amygdala neurons. J Neurosci 2015, 35:13020-13028.
- 53. Trouillet A-C, Keller M, Weiss J, Leinders-Zufall T, Birnbaumer L,
- Zufall F, Chamero P: Central role of G protein Gαi2 and Gαi2+ vomeronasal neurons in balancing territorial and infantdirected aggression of male mice. Proc Natl Acad Sci U S A 2019, 116:5135-5143

Trouillet *et al.* observe that conditional ablation of $G\alpha i2$ (which abolishes pheromonal responses in a subset of VNO neurons) enhances male-male aggression, but surprisingly decreases infant-directed aggression, with distinct neuronal activity profiles in downstream brain areas. Together with Ref. [37], this study provides evidence that these two types of aggression are controlled by different circuit mechanisms.

- Sheehan T, Paul M, Amaral E, Numan M, Numan M: Evidence that the medial amygdala projects to the anterior/ventromedial hypothalamic nuclei to inhibit maternal behavior in rats. *Neuroscience* 2001. 106:341-356.
- 55. Li Y, Mathis A, Grewe BF, Osterhout JA, Ahanonu B, Schnitzer MJ, Murthy VN, Dulac C: Neuronal representation of social information in the medial amygdala of awake behaving mice. *Cell* 2017, **171**:1176-1190.e17.
- 56. Tsuneoka Y, Tokita K, Yoshihara C, Amano T, Esposito G,
- Huang AJ, Yu LM, Odaka Y, Shinozuka K, McHugh TJ et al.: Distinct preoptic-BST nuclei dissociate paternal and infanticidal behavior in mice. EMBO J 2015, 34:2652-2670

In this careful study, Tsuneoka *et al.* observe characteristic and distinct patterns of neuronal activation in the central part of the MPOA (cMPOA) and the rhomboid nucleus of the BNST (BNSTrh) after paternal and infanticidal behavior, respectively. Intriguingly, cMPOA lesions switch fathers from paternal to infanticidal, whereas BNSTrh lesions inhibit infanticide in virgin males.

 57. Amano T, Shindo S, Yoshihara C, Tsuneoka Y, Uki H, Minami M,
 Kuroda KO: Development-dependent behavioral change toward pups and synaptic transmission in the rhomboid nucleus of the bed nucleus of the stria terminalis. Behav Brain Res 2017, 325:131-137

In this follow-up study to Ref. [56] (which reports that BNSTrh lesions suppress infanticidal behavior), the authors hypothesize that the transition from infant-directed affiliative behavior to aggression in males results from plastic changes in the BNSTrh. Using whole-cell patch clamp recordings, they find differences in evoked excitatory synaptic currents between adults and juveniles. This further highlights the BNSTrh as an important locus mediating the adaptive change from parental to infanticidal behavior in male mice.

 Renier N, Adams EL, Kirst C, Wu Z, Azevedo R, Kohl J, Autry AE, Kadiri L, Umadevi Venkataraju K, Zhou Y et al.: Mapping of brain activity by automated volume analysis of immediate early genes. *Cell* 2016, 165:1789-1802.

- Andermann ML, Lowell BB: Toward a wiring diagram understanding of appetite control. Neuron 2017, 95:757-778.
- Hashikawa K, Hashikawa Y, Falkner A, Lin D: The neural circuits of mating and fighting in male mice. Curr Opin Neurobiol 2016, 38:27-37.
- Chatterjee S, Sullivan HA, MacLennan BJ, Xu R, Hou Y, Lavin TK, Lea NE, Michalski JE, Babcock KR, Dietrich S et al.: Nontoxic, double-deletion-mutant rabies viral vectors for retrograde targeting of projection neurons. Nat Neurosci 2018, 21:638-646.
- Reardon TR, Murray AJ, Turi GF, Wirblich C, Croce KR, Schnell MJ, Jessell TM, Losonczy A: Rabies virus CVS-N2c ΔG strain enhances retrograde synaptic transfer and neuronal viability. Neuron 2016, 89:711-724.
- Ciabatti E, González-Rueda A, Mariotti L, Morgese F, Tripodi M: Life-long genetic and functional access to neural circuits using self-inactivating rabies virus. *Cell* 2017, 170:382-392.e14.
- Chan KY, Jang MJ, Yoo BB, Greenbaum A, Ravi N, Wu W-L, Sánchez-Guardado L, Lois C, Mazmanian SK, Deverman BE et al.: Engineered AAVs for efficient noninvasive gene delivery to the central and peripheral nervous systems. Nat Neurosci 2017, 20:1172-1179.
- 65. Tervo DGR, Hwang B-Y, Viswanathan S, Gaj T, Lavzin M, Ritola KD, Lindo S, Michael S, Kuleshova E, Ojala D et al.: A designer AAV variant permits efficient retrograde access to projection neurons. Neuron 2016, 92:372-382.
- Wang Y, Xie P, Gong H, Zhou Z, Kuang X, Wang Y, Li A, Li Y, Liu L, Veldman MB et al.: Complete single neuron reconstruction reveals morphological diversity in molecularly defined claustral and cortical neuron types. *bioRxiv* 2019:675280.
- 67. Winnubst J, Bas E, Ferreira TA, Wu Z, Economo MN, Edson P, Arthur BJ, Bruns C, Rokicki K, Schauder D *et al.*: Reconstruction of 1,000 projection neurons reveals new cell types and organization of long-range connectivity in the mouse brain. *bioRxiv* 2019:537233.
- Kebschull JM, Garcia da Silva P, Reid AP, Peikon ID, Albeanu DF, Zador AM: High-throughput mapping of single-neuron projections by sequencing of barcoded RNA. Neuron 2016, 91:975-987.
- Jun JJ, Steinmetz NA, Siegle JH, Denman DJ, Bauza M, Barbarits B, Lee AK, Anastassiou CA, Andrei A, Aydn Ç et al.: Fully integrated silicon probes for high-density recording of neural activity. Nature 2017, 551:232-236.
- Allen WE, Chen MZ, Pichamoorthy N, Tien RH, Pachitariu M, Luo L, Deisseroth K: Thirst regulates motivated behavior through modulation of brainwide neural population dynamics. Science 2019, 364:253.
- Stringer C, Pachitariu M, Steinmetz N, Reddy CB, Carandini M, Harris KD: Spontaneous behaviors drive multidimensional, brainwide activity. Science (80-) 2019, 364:eaav7893.
- Stringer C, Pachitariu M, Steinmetz N, Carandini M, Harris KD: High-dimensional geometry of population responses in visual cortex. *Nature* 2019, 571:361-365.
- Mathis A, Mamidanna P, Cury KM, Abe T, Murthy VN, Mathis MW, Bethge M: DeepLabCut: markerless pose estimation of userdefined body parts with deep learning. Nat Neurosci 2018, 21:1281-1289.
- 74. Graving JM, Chae D, Naik H, Li L, Koger B, Costelloe BR, Couzin ID: DeepPoseKit, a software toolkit for fast and robust animal pose estimation using deep learning. *eLife* 2019, 8.
- Hong W, Kennedy A, Burgos-Artizzu XP, Zelikowsky M, Navonne SG, Perona P, Anderson DJ: Automated measurement of mouse social behaviors using depth sensing, video tracking, and machine learning. Proc Natl Acad Sci U S A 2015, 12:E5351-E5360.